

Zinc-containing enzyme; activation of β - and γ -carbonic anhydrases from pathogenic bacteria with tripeptides

Marilisa Pia Dimmito^a, Azzurra Stefanucci^a, Adriano Mollica^a

^aDepartment of Pharmacy, "Gabriele d'Annunzio" University of Chieti-Pescara, Chieti, Italy;

Topic: Old Elements, New Technologies: how to improve the quality of life.

Abstract:

Zinc is a chemical element with atomic number 30, turns out to be one of the essential elements present in living organisms. Zinc is located in the active site of Carbonic Anhydrases (CA) and is responsible of its catalytic mechanism. To date, several isoforms of anhydrase are known, in particular α - β - γ - Carbonic Anhydrases are Zinc dependent and are very important in medicinal chemistry for their catalytic activity towards CO_2 .¹

In living organisms, the hydration of CO_2 and the dehydration of HCO_3^- take place in a very fast process, like all processes connected to transport / secretion. The main metabolic role of CA is the high efficiency catalysis of CO_2 hydration (**Figure 1**).

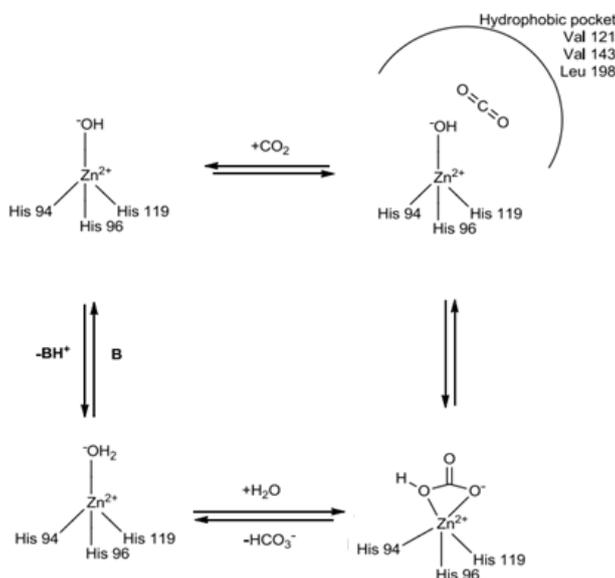


Figure 1. Interconversion between carbon dioxide and bicarbonate.

CA inhibitors (CAIs) show pharmacologic applications in pathologies in which the activity of these enzymes is dysregulated (in humans), such as edema, glaucoma, neurologic diseases, obesity, and some tumors, and many sulphonamide or sulphamate CAIs have been in clinical use for decades.²

In contrast, investigation of activators of these enzymes (CAAs) have been relatively limited.³

However, no drug-design CA studies are available for bacterial, β - and γ -CAs. These enzymes have only recently been started to be investigated for their activation with amines and amino acids. Bacterial β -CAs are active in both dimeric and tetrameric forms, with two or four identical active sites. Their closed conformation structure represents the catalytic Zinc ion placed in the terminal end of a channel, coordinated tetrahedral with two cysteines, an histidine and an aspartic acid residue. The structure of the enzymes synthesized by *Vibrio cholera* ($VchCA\beta$) is particular since its catalytic active site in the closed form, has been isolated at pH lower than 8.3 (**Figure 2**). In the inactive form the bicarbonate establishes bonds with the Zinc ion inside the closed pocket, however at pH > 8.3 the closed active site assumes the open conformation because a residue of Asp, near the metal ion, moves to discover the Zinc ion which can coordinate with a water molecule involved in catalytic activity.⁴

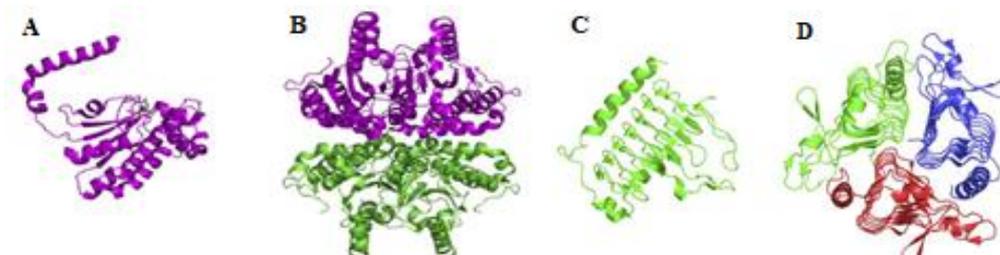


Figura 2 Representation of **A** inactive monomer, **B** active tetramer of β -CA of *Vibrio cholera*; **C** inactive and **D** active trimer of the γ -CA enzyme coming from the bacterium *Methanosarcina thermophila*.

β -CA adopts a 3-D conformation β -helix and crystallizes as a tetramer with three active sites containing Zinc, each of which is at the interface between two monomers. The enzyme metal is active only in trimeric form.⁵ This enzymatic class presents a residue of glutamic acid in place of a histidine, which acts as a proton transporter. The design of bioactive molecules that modulate these enzymes may be useful for controlling the intra and extracellular pH of microorganisms, which can play crucial roles in the life cycles of pathogenic microorganisms. This is of great interest especially to counter the problem associated with the increase of resistance that microorganisms present towards conventional antibiotics. In this work we report a study for investigating the capability of tripeptides incorporating amino acid residues do show activating effects against β - and γ - class CAs from pathogenic bacteria such as *Vibrio Cholerae*, *Mycobacterium tuberculosis* and *Burkholderia pseudomallei*. These tripeptides may be useful tools for investigating the role of these enzymes in key bacterial processes such as invasion, colonization and pathogenicity, which are currently poorly understood.⁶

References

1. C.T. Supuran. *Biochem. J.* **2016**, 473, 2023–32.
2. C.T. Supuran. *Expert Opin Drug Discov* **2017**, 12, 61–88.
3. C.T. Supuran. *Nat Rev Drug Discov* **2008**, 7, 168–81.
4. S. Huang, T. Hainzl, C. Grundstrom, C. Forsman, G. Samuelsson, A. E. Sauer-Eriksson. *PLoS One* **2011**, 6, E28458.
5. B.C. Tripp, J. G. Ferry. *Biochemistry* **2000**, 39, 9232–40.
6. A. Stefanucci, A. Angeli, M.P. Dimmito, G. Luisi, S. Del Prete, C. Capasso, W.A. Donald, A. Mollica & C. T. Supuran. *J. of Enz. Inhib. and Med. Chem.* **2018**, 33, 945–950.